

Biosynthesis of the Nicandrenoids: Stages in the Oxidative Elaboration of the Side Chain and the Fate of the Diastereotopic 25-Methyl Groups of 24-Methylenecholesterol

Harjit K. Gill, Roland W. Smith, and Donald A. Whiting*

Department of Chemistry, The University, Nottingham NG7 2RD, U.K.

Experiments with *Nicandra physaloides* plants show that the insect antifeedant steroid Nic-1 (**1**) is formed from 24-methylenecholesterol: in double bond isomerisation to 24-methylcholesta-5,24-dien-3 β -ol, the 25-(*pro-S*) methyl in (**2**) becomes the 25-(*pro-Z*) methyl, in (**3**); further oxidations lead to triol (**6**) and hence to lactol (**1**) with partial retention of 26-hydrogen.

The withanolides,¹ a large group of 24-methyl plant steroids occurring in the Solanaceae, are characterised by extensive oxidative modifications. The insect antifeedant Nic-1 (**1**), belonging to a distinctive sub-set isolated from *Nicandra physaloides*,² contains, as well as the unique aromatic ring-D,³ an unusual epoxy lactol side chain. This feature also appears in Nic-3 and, in modified form, in other nicandrenoids. Structural comparisons within the group^{1,4} suggest that side chain modification may either precede ring A/B modification or be partly independent of the A/B system (as in a metabolic grid with enzymes of low substrate specificity). We report here experiments which confirm this view and outline stages in the oxidative development of Nic-1: selectivity in the oxidation of the diastereotopic C-25 methyl groups in 24-methylenecholesterol (**2**) is revealed in the sequence.

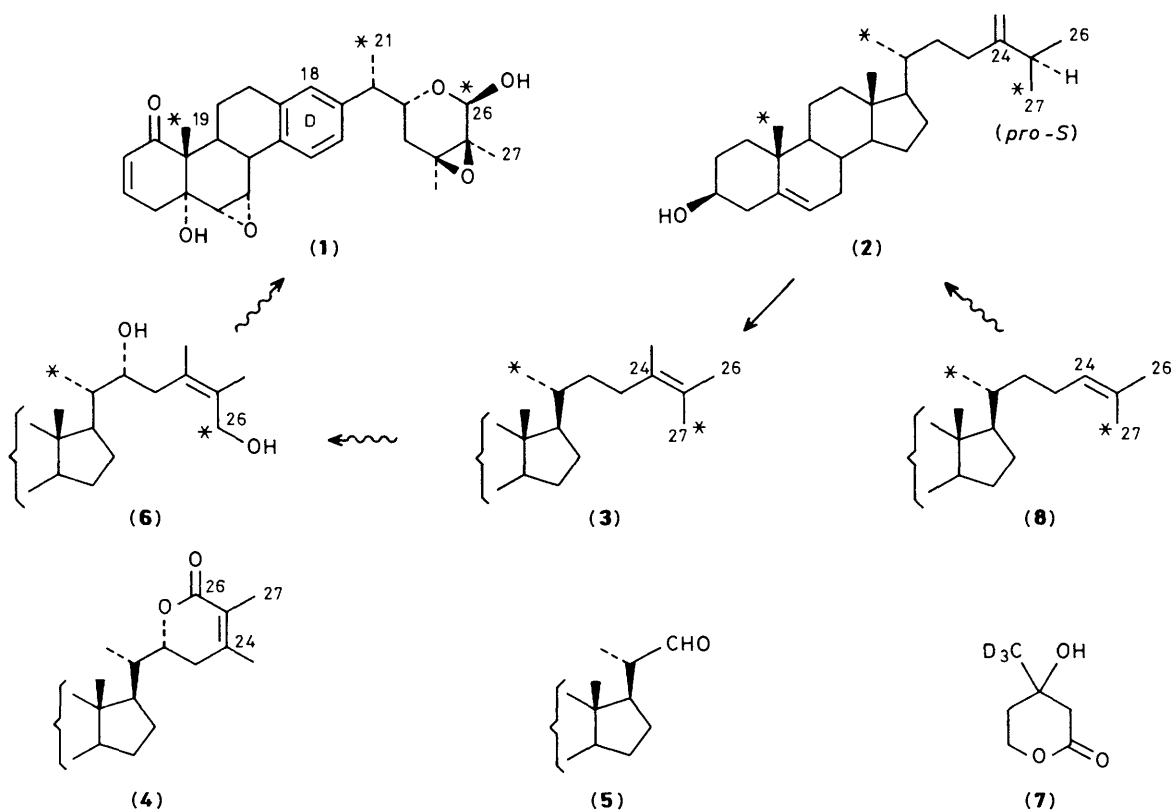
As potential precursors to Nic-1 we have prepared (i) [28-¹⁴C]-24-methylenecholesterol (**2**), from 24-oxocholesterol;⁵ (ii) [28-¹⁴C]-24-methylcholesta-5,24-dien-3 β -ol (**3**), by isomerisation of (**2**);⁶ (iii) the [23,28-³H₂]-(*22R*)-lactone (**4**), synthesised from aldehyde (**5**)⁷ using methods similar to those of Glotter and coworkers,⁸ and labelled by tritium

exchange (³H₂O-1,5-diazabicyclo[4.3.0]non-5-ene); and (iv) the [23,28-³H₂]-(*22R*),(*24Z*)-diol (**6**), by LiAlH₄ reduction of lactone (**4**). The four sterols were then administered to seven week old *Nicandra physaloides* plants in Tween-20-water-2-methoxyethanol, through cut stems. After 4 days metabolism, Nic-1 was isolated and recrystallised to constant activity; the incorporations, based on sterol uptake, are shown in Table 1.

The results indicate that both 24-methylenecholesterol, the first product from *S*-adenosylmethionine methylation of

Table 1. Absolute (%) and specific (%) incorporations of labelled sterols into Nic-1 in *N. physaloides* plants.

	Absolute incorporation	Specific incorporation
(2)	0.23	0.056
(3)	0.16	0.068
(4)	0.17	0.015
(6)	0.50	0.057



cholesterol, and its 24(25)-double bond isomer (3) are incorporated, at levels expected in such experiments: these two sterols are probably adjacent in the biosynthetic sequence. The triol (6) shows a higher absolute incorporation, as appropriate to a later stage precursor and is a distinctly better utilised precursor than the lactone (4). This suggests that the predominant pathway from alcohol (6) to the hemiacetal (1) proceeds by way of a C-26 aldehyde, in preference to reduction of the lactone (4) (although the latter route may constitute a minor pathway). This view is reinforced by the outcome of administration of [$3'$ - $C^{14}H_3$]-mevalonolactone (7) to *N. physaloides*. Examination of the 2H n.m.r. spectrum of the resulting Nic-1 shows signals arising from (i) the methyl groups (C-19, C-21), as expected; (ii) 18-H, see ref. 3, and (iii) 26-H (δ 4.9). This indicates that C-26 in Nic-1 is derived from the mevalonic acid 3'-methyl with a degree of retention of hydrogen, thus excluding lactone (4), or a related compound with C-26 fully oxidised, from the predominant pathway.

A further inference may be made from this experiment. It has been shown⁹ that, in the biosynthesis of 24-methylenecholesterol in a closely related solanaceous plant, *Physalis peruviana*, C-27 of lanosterol† (8) becomes C-27 [the 25-(*pro-S*)-methyl] of 24-methylenecholesterol (2). Since C-26 of Nic-1 must derive from C-27 of sterol (3) then it follows that

this latter carbon must correlate with C-27 of sterol (2), so that the *net* methylation of lanosterol† (8) to sterol (3) proceeds with retention of double bond geometry.

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† A closely related alternative precursor may be involved.